



## Analytical Procedures

July 2015

### Ash

AOAC Method 942.05 – Ash of Animal Feed.

### Carbohydrates

#### **Ethanol Soluble Carbohydrates (ESC)**

Hall, M.B., W.H. Hoover, J.P. Jennings and T.K. Miller Webster. 1999. A method for partitioning neutral detergent soluble carbohydrates. *J. Sci. Food Agric.* 79: p.2079-2086.

Samples shaken for 4 hours at 180 epm with 80% ethanol to extract ethanol soluble carbohydrates comprised of simple sugars. ESC determined using a Thermo Scientific Genesys 10S Vis Spectrophotometer after a colorimetric phenol-sulfuric acid reaction.

#### **Water Soluble Carbohydrates (WSC)**

West Virginia University Procedure by W.H. Hoover and T.K. Miller Webster. Determination of Nonstructural Carbohydrates.

Hall, M.B., W.H. Hoover, J.P. Jennings and T.K. Miller Webster. 1999. A method for partitioning neutral detergent soluble carbohydrates. *J. Sci. Food Agric.* 79: p.2081.

Samples incubated with water in a 40°C bath for 1 hour extracting water soluble carbohydrates comprised of simple sugars and fructan. WSC determined using a Thermo Scientific Genesys 10S Vis Spectrophotometer after acid hydrolysis with sulfuric acid and colorimetric reaction with potassium ferricyanide.

### Carbon (C)

Dry, 1mm ground samples analyzed using a Leco CN628 Carbon/Nitrogen Determinator.  
Leco Corporation, 300 Lakeview Avenue, St. Joseph, MI 49085. [www.leco.com](http://www.leco.com)

### Coliform and *E. coli*, water

Colilert® - IDEXX Laboratories, Inc., One IDEXX Drive, Westbrook, Maine 04092. [www.idexx.com](http://www.idexx.com)

Colilert® is a presence/absence test that detects total coliforms and *E. coli* at 1 organism/100 ml that uses the patented Defined Substrate Technology® (DST®) to simultaneously detect total coliforms and *E. coli*. Two nutrient-indicators, ONPG and MUG, are the major sources of carbon in Colilert® and can be metabolized by the coliform enzyme  $\beta$ -galactosidase and the *E. coli* enzyme  $\beta$ -glucuronidase, respectively.

As coliforms grow in Colilert®, they use  $\beta$ -galactosidase to metabolize ONPG and change it from colorless to yellow to indicate presence. *E. coli* use  $\beta$ -glucuronidase to metabolize MUG and create fluorescence to indicate presence. Since most non-coliforms do not have these enzymes, they are unable to grow and interfere.

Colilert® is US FDA Approved for Dairy Waters.

- Milk Laboratory Evaluation Form FDA 2400m (3/01)

Colilert® is also US EPA-approved for drinking water presence/absence (P/A) and Most Probable Number (MPN) and for source water. Pertinent references:

- June 29, 1989 US EPA Federal Register Colilert® coliform approval
- June 10, 1992 US EPA Federal Register Colilert® *E. coli* approval

## Corn Stalk Nitrogen Testing (CSNT)

Miller, R.O. 1998. Extractable nitrate in plant tissue: ion selective method. p85-88. Y.P. Kalra (ed.) Handbook of Reference Methods for Plant analysis. CRC Press LLC, Boca Raton, FL.

Wilhelm, W.W., S.L. Arnold, and J.S. Schepers. 2000. Using nitrate specific ion electrode to determine stalk nitrate-nitrogen concentration. Agron. J. 92:186-189.

Wilhelm, W.W., G.E. Varvel, and J.S. Schepers. 2005. Corn stalk nitrate concentration profile. Agron. J. 97:1502-1507.

Stalk Analysis. Methodology prepared by Soil Nutrient Laboratory, University of Connecticut.

Stalk Analysis. Methodology prepared by Cornell Nutrient Analysis Laboratory, Cornell University.

## Density, manure

Standard Vial Method. 2002. Dairy One. Samples weighed into fixed volume vessel. Density calculated and expressed in kg/l, lbs./ft<sup>3</sup>, and lbs./gal.

## Dry Matter (DM)

### Oven – 60°C for 4 hours (forced air)

Goering, H.K. and P.J. Van Soest. 1970. Forage Fiber Analyses (apparatus, reagents, procedures, and some applications). ARS/USDA Handbook No. 379, Superintendent of Documents, US Government Printing Office, Washington, D.C. 20402. P15.

NFTA Method 2.2.1.1 – Partial Dry Matter using Forced-air Drying Ovens.

### Oven – 135°C for 2 hours

AOAC 930.15 – Loss on Drying (Moisture) for Feeds.

### Oven – 105°C for 3 hours

NFTA Method 2.2.2.5 – Dry Matter by Oven Drying for 3hr at 105C.

## Fat

### Crude, Acid Hydrolysis

AOAC 954.02 – Crude Fat in Pet Food.

### Crude, Ether Extraction

AOAC 2003.05 – Crude Fat in Feeds, Cereal Grains, and Forages.

Extraction by Soxtec HT6 System using anhydrous diethyl ether. Crude fat residue determined gravimetrically after drying.

Foss North America, 8091 Wallace Road, Eden Prairie, MN 55344. [www.foss.us](http://www.foss.us)

### Crude, Roese-Gottlieb Method

AOAC 932.06 A (b) and 932.06 B – Fat in Dried Milk.

## Fiber

### Acid Detergent Fiber (ADF)

ANKOM Technology Method 5 – Acid Detergent Fiber in Feeds – Filter Bag Technique for A200 (4-13-11).

Solutions as in AOAC 973.18 – Fiber (Acid Detergent) and Lignin (H<sub>2</sub>SO<sub>4</sub>) in Animal Feed. Samples individually weighed at 0.5g into filter bags and digested for 75 minutes as a group of 24 in 2L of ADF solution in ANKOM A200 Digestion Unit. Samples are rinsed three times with boiling water for 5 minutes in filter bags followed by a 3 minute acetone soak and drying at 105°C for 2 hours.

ANKOM Technology, 2052 O'Neil Road, Macedon, NY 14502. [www.ankom.com](http://www.ankom.com)

## Fiber (continued)

### Crude Fiber

ANKOM Technology Method 1 – Crude Fiber Analysis in Feeds – Filter Bag Technique (FBT) – AOAC Approved Procedure Ba 6a-05 (Rev E 4-13-11).

Same digestion solutions as in AOAC 962.09 – Fiber (Crude) in Animal Feed and Pet Food. Acetone used for presoak fat extraction. 0.5g weighed for forages, dried manure, wood, and wood pellets. 1.0g weighed for all other sample types.

### Lignin

ANKOM Technology Method 9 – Method for Determining Acid Detergent Lignin in the Daisy<sup>II</sup> Incubator – 04/11.

Solution as in AOAC 973.18 – Fiber (Acid Detergent) and Lignin (H<sub>2</sub>SO<sub>4</sub>) in Animal Feed. ADF performed as above and residue digested as a group of 24 in 72% w/w sulfuric acid for 3 hours in ANKOM Daisy<sup>II</sup> Incubator at ambient temperature.

### aNDF (amylase and sodium sulfite treated Neutral Detergent Fiber)

ANKOM Technology Method 6 – Neutral Detergent Fiber in Feeds – Filter Bag Technique for A200 (4-13-11).

Solutions as in Van Soest, P.J., J.B. Robertson, and B.A. Lewis. 1991. Methods for Dietary Fiber, Neutral Detergent Fiber, and Nonstarch Polysaccharides in Relation to Animal Nutrition. *J.Dairy Science* 74:3583-3597.

Samples individually weighed at 0.5g into filter bags and digested for 75 minutes as a group of 24 in 2L of NDF solution in ANKOM A200 Digestion Unit. Four ml of Alpha Amylase and 20g sodium sulfite are added at the start of digestion. Samples are rinsed three times with boiling water for 5 minutes. Alpha Amylase is added to the first 2 rinses. Water rinses are followed by a 3 minute acetone soak and drying at 105°C for 2 hours.

### aNDFom (aNDF on an organic matter (ash free) basis)

aNDF analyzed as above but with the addition of an ashing step to remove inorganic materials such as minerals, soil, and sand by burning the fibrous residue at 550C for 2 hours.

### NDFD (Neutral Detergent Fiber Digestibility – 24, 30, 48 hr. time points)

ANKOM Technology Method 3 – In Vitro True Digestibility using the Daisy<sup>II</sup> Incubator (08/05).

Reagents and solutions as in: Goering, H.K. and P.J. Van Soest. 1970. Forage Fiber Analyses (apparatus, reagents, procedures, and some applications). ARS/USDA Handbook No. 379, Superintendent of Documents, US Government Printing Office, Washington, D.C. 20402. P13-14.

Rumen fluid collected from TMR fed, high producing lactating cow. Dry, ground feed samples (0.25g; 1mm particle size) incubated in Van Soest buffer/rumen fluid mixture for specified hours under anaerobic conditions at 39°C. After incubation, samples extracted using aNDF procedure to remove bacterial contamination. Residue is undigested fibrous material and is used to determine in-vitro true digestibility (IVTD) and neutral detergent fiber digestibility (NDFD). Lag time 4.9 hours.

### NDFDom (Neutral Detergent Fiber Digestibility on an organic matter (ash free) basis – 12, 72, 120 or 30, 120, 240 hrs.)

NDF Digestibility (NDFD) analyzed as above but with the addition of an ashing step to remove inorganic materials such as minerals, soil, and sand by burning the undigested fibrous residue at 550C for 2 hours. NDFDom results expressed on an organic matter (ash free) basis as a percentage of the aNDFom.

### uNDFom (undigestible Neutral Detergent Fiber on organic matter (ash free) basis – 12, 72, 120 or 30, 120, 240 hrs.)

NDF Digestibility (NDFD) analyzed as above but with the addition of an ashing step to remove inorganic materials such as minerals, soil, and sand by burning the undigested fibrous residue at 550C for 2 hours. Undigested NDF expressed on an organic matter (ash free) basis as a percentage of the dry matter.

## Gross energy (GE)

Gross energy (gross calorific value) of solid and liquid materials expressed as calories per gram (cal/g) using an IKA C2000 basic Calorimeter System.

Instrument is set to IKA's dynamic mode with an outer vessel temperature set at 25°C. Analysis time is 7-12 minutes. Dried samples weighed into polyethylene bags. Oil type samples weighed into gelatin capsules. Samples placed in a crucible, then ignited in an oxygen rich atmosphere in a sealed decomposition vessel where the increase in temperature of the system is measured.

The specific gross calorific value of the sample is calculated from:

- The weight of the sample.
- The heat capacity of the calorimeter system determined from benzoic acid calibration standards.
- The increase in temperature of the water within the inner vessel of the measuring cell.

IKA Works, Inc., 2635 North Chase Pkwy SE, Wilmington, NC 28405-7419. [www.ika.com](http://www.ika.com)

## Minerals

### Ca, P, Mg, K, Na, Fe, Zn, Cu, Mn, Mo, Co, S, Al, B, Cr, Sr

Samples digested using CEM Microwave Accelerated Reaction System (MARS6) with MarsXpress Temperature Control using 50ml calibrated Xpress Teflon PFA vessels with Kevlar/fiberglass insulating sleeves then analyzed by ICP using a Thermo iCAP 6300 Inductively Coupled Plasma Radial Spectrometer.

Sample weights – 0.5g for forages, ingredients, byproducts (1.0g for Co or Cr); 0.5g for grain mixes; 0.2g for mineral mixes; Manure - 0.5g dried, ground or 2-10g wet sample.

Samples first pre-digested at ambient temperature 10 minutes with 8ml nitric acid (HNO<sub>3</sub>) and 2ml hydrochloric acid (HCl) and then an additional 10 minutes with 1ml 30% hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). After pre-digestion complete, samples ramped to 200°C in 15 minutes and finally held at digestion temperature of 200°C for 15 minutes at 1600W. Vessels brought to 50-ml volume, aliquot used for analysis.

Method utilized based upon CEM Application Notes for Acid Digestion on the following matrices - Feed Grain, Alfalfa, Corn Flour, Milk Powder, Soybean Meal, Flour, Hair, Potato Chips, Wheat Crackers, Peanut Butter, Urine, Dog Feces, Wine.

Water – 35ul concentrated nitric acid added to 14ml of water, mixed, then aspirated on ICP for analysis.

Manure Reference: Wolf, Ann, M. Watson, and N. Wolf. 2003. Digestion and dissolution methods for P, K, Ca, Mg and trace elements. Recommended methods of manure analysis. ed J. Peters, pp30-39. University of Wisconsin Extension Publication. A3769

CEM, 3100 Smith Farm Road, Matthews, NC 28106. [www.cem.com](http://www.cem.com)  
Thermo Fisher Scientific Inc., 81 Wyman Street, Waltham, MA 02454. [www.thermoscientific.com](http://www.thermoscientific.com)

### Chloride Ion (Cl<sup>-</sup>)

Brinkmann Metrohm 716 Titrimetric Unit - 0.5g dried, ground sample or 5g wet sample extracted for 15 minutes in 50ml 0.1N HNO<sub>3</sub>, followed by potentiometric titration with AgNO<sub>3</sub> using Brinkmann Metrohm 716 Titrimetric Unit with silver electrode. For water samples, 25ml of 0.2N HNO<sub>3</sub> added to 25ml of sample then analyzed.

Metrohm Application Bulletin No. 130 by Metrohm Ltd., C-H-9101 Herisau, Switzerland  
Metrohm USA, 6555 Pelican Creek Circle, Riverview FL, 33578. [www.metrohmusa.com](http://www.metrohmusa.com)

The method by Metrohm is similar to the concepts found in: Cantliffe, D.J., MacDonald, G.E. and Peck, N.H. 1970. The potentiometric determination of nitrate and chloride in plant tissue. New York's Food and Life Sciences Bulletin. No.3, September 1970. Plant Sciences. Vegetable Crops Geneva. No. 1: 5-7.

## Near Infrared Reflectance Spectroscopy (NIRS)

AOAC 991.01 – Moisture in Forage.

AOAC 989.03 – Fiber (Acid Detergent) and Protein (Crude) in Forages.

Foss NIRSystems Model 6500 with Win ISI II v1.5. Components analyzed by NIRS include: DM, CP, SP, RDP, ADI-CP, NDI-CP, ADF, aNDF, lignin, starch, WSC, ESC, fat, ash, Ca, P, Mg, K, S, Cl, NDFD 24, NDFD 30, NDFD 48, starch digestibility, aNDFom, uNDFom 30, 120, 240, NDFDom 30, 120, 240. Silages also receive lactic acid, acetic acid, and ammonia CPE.

Near infrared reflectance spectroscopy (NIR) is an instrumental method for rapidly and reproducibly measuring the chemical composition of forage and feed samples. It is based on the fact that each of the major chemical components of a sample has near infrared absorption properties that can be used to 1) differentiate one component from another and 2) determine nutrient concentration.

NIR is a calibration based technology, meaning that analysis is limited to only feeds and nutrients for which calibrations have been developed. Dairy One has built broad based calibrations by incorporating samples collected over several decades. Calibrations are based on reference chemistry using traditional procedures. Visit this link for more info <http://dairyone.com/analytical-services/feed-and-forage/services-and-pricing/>.

Foss NIRSystems, 7703 Montpelier Road, Suite 1, Laurel, MD 20723. [www.foss.us](http://www.foss.us)

## Nitrates (%NO<sub>3</sub> or ppm NO<sub>3</sub>-N)

RQflex<sup>®</sup> Reflectometer Method

1g of dried, ground sample or 10g of wet sample is extracted in 50ml deionized water for 20 minutes by shaking at 280 oscillations/minute. Samples are filtered through Whatman 934-AH (1.5µm) filter paper, then analyzed by RQflex<sup>®</sup> Reflectometer using Reflectoquant<sup>®</sup> Nitrate test strips.

When the Nitrate test strip is immersed in the aqueous sample, a reducing agent reduces nitrate ions to nitrite ions. In the presence of an acidic buffer, the nitrite ions react with an aromatic amine to form a diazonium salt. The salt reacts with N-(1-naphthyl)-ethylene-diamine to form a red-violet azo dye that is measured reflectometrically. Nitrate concentration is proportional to the color reaction.

Each strip contains two reaction zones generating dual replicate analyses per sample. The RQflex<sup>®</sup> Reflectometer's double optic system measures the analyte concentration based on the light reflected from the dual reaction zones. Barcode-controlled software calculates the mean of those two measurements.

EMD Chemicals Inc., One International Plaza, Suite 300, Philadelphia, PA, 19113. [www.emdmillipore.com](http://www.emdmillipore.com)

## pH

Feed and Forage -15g wet sample placed into 250-ml beaker. 200ml deionized water added, stirred, and allowed to stabilize five minutes. Manure -35ml liquid sample poured into 50ml beaker. 15g solid or semi-solid sample weighed into 200 ml deionized water, stirred, and allowed to stabilize five minutes. Water - AOAC 973.41 – pH of Water.

Analyzed using Thermo Orion Combination Sure-Flow pH Electrode and Thermo Orion 410 A meter. Calibrated with buffers referenced to NIST SRMs. pH 4 buffer contains potassium hydrogen phthalate and pH 7 buffer contains sodium phosphate dibasic and potassium phosphate monobasic.

Thermo Fisher Scientific, 81 Wyman Street, Waltham, MA 02454. [www.thermoscientific.com](http://www.thermoscientific.com)

## Protein

### Acid Detergent Insoluble Crude Protein (ADICP)

ADF residue analyzed using a Leco TruMac N Macro Determinator to determine the protein fraction bound to the acid detergent fiber.

### Crude Protein (CP) and Total Nitrogen (N)

Dry, 1mm ground samples analyzed by combustion using a CN628 Carbon/Nitrogen Determinator. Liquid samples analyzed using a TruMac N Macro Determinator.

AOAC 990.03 – Protein (Crude) in Animal Feed

AOAC 992.15 – Crude Protein in Meat and Meat Products including Pet Foods

AOAC 992.23 – Crude Protein in Cereal Grain and Oilseeds

Leco Application Note – “Nitrogen/Protein in Feeds, Grains, and Oil Seeds” Form 203-821-392, 09/10-Rev0.

Leco Application Note – “Nitrogen in Soil and Plant Tissue” Form 203-821-394, 09/10-Rev0.

Manure – Watson, M., A. Wolf, and N. Wolf. 2003. Total nitrogen. Recommended methods of manure analysis. ed J. Peters, pp18, 23-24. University of Wisconsin Extension Publication. A3769.

Leco Corporation, 300 Lakeview Avenue, St. Joseph, MI 49085. [www.leco.com](http://www.leco.com)

### Degradable Protein (Rumen Degradable Protein - RDP)

Cornell *Streptomyces griseus* (SGP) enzymatic digestion. Enzyme concentration held constant. Residues containing undegradable protein analyzed using Leco TruMac N Macro Determinator.

Concentrates incubated for 18 hrs. Cornell Nutrition Conference Proceedings, 1990. pp. 81-88.

Forage samples incubated for 2 hrs at higher SGP concentration. J. Dairy Sci. 1999. 82: 343-354.

### Neutral Detergent Insoluble Crude Protein (NDICP)

aNDF performed without sodium sulfite then residue analyzed using a Leco TruMac N Macro Determinator to determine the protein fraction bound to the neutral detergent fiber.

## Protein (continued)

### Non Protein Nitrogen (NPN)

#### Ammonia Crude Protein Equivalent (CPE) or Ammonium-N

Timberline TL-2800 Analyzer

Extraction - Samples are extracted in deionized water using a single speed blender at 20,000 rpm for 2 minutes (50g/750ml) or a reciprocal shaker for 30 minutes at 280 epm (Forage - 5g/100ml wet or 1g/100ml dry; Manure - 10g/150ml). For urea, a prepared urease solution is added to a duplicate sample prior to shaking (5g/100ml wet or 1g/100ml dry). All extracts then centrifuged at 4000 rpm for 5 minutes, decanted into tubes, then analyzed.

Analysis - A peristaltic pump directs the sample, caustic, and absorbing solutions into a diffusion cell. Within the cell, the sample is mixed with the caustic solution, resulting in a pH of 11-13 which converts the ammonium ion present in the sample to dissolved ammonia gas. The sample/caustic solution flows past one side of a membrane that is permeable to gases but not to liquids or ionic species. The dissolved ammonia gas in the sample/caustic mixture diffuses across the membrane. On the other side of the membrane, a buffered solution absorbs the diffused ammonia gas then flows through a low volume heat exchanger to establish thermal equilibrium then into the conductivity detector. The conductivity cell measures the change in electrical conductance of the absorbing solution. This change is proportional to the concentration of ammonium in the original sample.

Timberline Instruments, 1880 S. Flatiron Ct. Suite I, Boulder, CO 80301. [www.timberlineinstruments.com](http://www.timberlineinstruments.com)

Extraction using reciprocating shaker – Kalra, Y.P. 1998. Determination of Ammonium-Nitrogen in Plant Tissue. *Handbook of Reference Methods for Plant Analysis*. 11:90.

Principles of operation – Carlson, R.M. 1978. Automated separation and conductimetric determination of ammonia and dissolved carbon dioxide. *Analytical Chemistry* 50:1528-1531.

#### Urea Crude Protein Equivalent (CPE)

Timberline TL-2800 Analyzer. Analyzed as above in Ammonia CPE after addition of prepared urease enzyme solution.

#### Urea Crude Protein Equivalent (CPE)

AOAC 967.07 – Urea in Animal Feed, colorimetric method. Alternate urea method used for samples containing high levels of minerals including mineral mixes, liquid supplements, molasses, and grain mixes with added minerals.

### Organic Nitrogen, manure

Calculated by difference (Total Nitrogen minus Ammonium-Nitrogen).

### Soluble Protein (SP)

Cornell Sodium Borate-Sodium Phosphate Buffer Procedure. Soy products incubated at 39°C. All other samples incubated at ambient temperature. Residue containing insoluble protein analyzed using Leco TruMac N Macro Determinator.

Cornell Nutrition Conference Proceedings, 1990, pp. 85-86.

## Standard Plate Count, water

US FDA Milk Laboratory Evaluation Form FDA 2400a (1/01)

Petrifilm Aerobic Count Method – 1ml of sample deposited onto petrifilm and covered. Sample distributed with spreader and gel allowed to solidify for 1 minute. Incubated 48 hours at 32°C. Colonies counted when incubation time is complete and reported as colonies per ml.

## Starch

YSI 2700 SELECT Biochemistry Analyzer

YSI Incorporated Life Sciences, 1725 Brannum Lane, Yellow Springs, Ohio 45387 Application Note Number 319.

[www.ysilifescience.com](http://www.ysilifescience.com)

Samples are pre-extracted for sugar by incubation in 40°C water bath and filtration on Whatman 41 filter paper. Residues are thermally solubilized using an autoclave, then incubated with glucoamylase enzyme to hydrolyze starch to produce dextrose (glucose).

Prepared samples injected into sample chamber of YSI Analyzer where dextrose diffuses into a membrane containing glucose oxidase. The dextrose is immediately oxidized to hydrogen peroxide and D-glucono-4-lactone. The hydrogen peroxide is detected amperometrically at the platinum electrode surface. The current flow at the electrode is directly proportional to the hydrogen peroxide concentration, and hence to the dextrose concentration. Starch is determined by multiplying dextrose by 0.9.

### **Sulfates, water (SO<sub>4</sub> and SO<sub>4</sub>-S)**

Turbidimetric Method. AQUAfast AC2082 Tablet Chemistry  
Thermo Fisher Scientific, 166 Cummings Center, Beverly, MA 01915, 1-800-225-1480. [www.thermo.com/water](http://www.thermo.com/water)

BaCl<sub>2</sub> tablet is added to the water sample resulting in precipitation of sulfate as BaSO<sub>4</sub>. Suspension is measured photometrically at 520nm with a Thermo Scientific AQUAfast AQ4000 Colorimeter to determine the sulfate concentration in mg/l (ppm). Sulfate-sulfur (SO<sub>4</sub>-S) calculated as sulfates (SO<sub>4</sub>) divided by 2.996.

### **Total Dissolved Solids, water (TDS)**

Conductivity Method. ES&D Model 76 Conductivity meter.

The total quantity of free ions is determined by ability of the sample to conduct an electrical current. Electrode immersed in water while gently stirring. Temperature of water measured, temperature knob set, meter allowed to stabilize for 15 seconds, and then reading recorded.

Engineered Systems & Designs, 119A Sandy Drive, Newark DE, 19713. [www.esdinc.com](http://www.esdinc.com)

### **Total Solids, manure**

#### **Oven – 105°C for 16 hours (gravity)**

Used for liquid or solid samples with no bedding.

#### **Oven – 60°C for 6-8 hours (forced air) + NIRS - AOAC 991.01 –Moisture in Forage.**

Used for Liquid or solid sample with bedding.

Hoskins, B., A. Wolf, and N. Wolf. 2003. Dry matter analysis. Recommended methods of manure analysis. ed J. Peters, pp14-17. University of Wisconsin Extension Publication. A3769.

### **Volatile Fatty Acids and Lactic Acid**

Extraction – 50g samples blended at 20000 rpm for 2 min. in 750ml deionized water (Manure 50g and 450ml water), filtered through cheesecloth, then filtered through disposable syringe filter. Adapted from Personal Communication, L.E. Chase, Ph.D., Cornell University.

#### **Gas Chromatography – Acetic, Propionic, Butyric, Iso-butyric acids**

Aliquot of extract mixed 1:1 ratio with 0.06M oxalic acid containing 100ppm trimethylacetic acid (internal standard). Samples injected into a Perkin Elmer Autosystem XL Gas Chromatograph containing a Supelco packed column with the following specifications: 2m x 2mm Tightspec ID, 4% Carbowax 20M phase on 80/120 Carbopack B-DA.

Procedure based upon:

- "GC Separation of VFA C2-C5" Supelco GC Bulletin 749F, 1975.
- "Analyzing Fatty Acids by Packed Column Gas Chromatography" Supelco GC Bulletin 856A, 1990.
- "Volatile Fatty Acid SOP" W.H. Miner Institute, Chazy, NY.

Sigma Aldrich (Supelco), 3050 Spruce Street, St. Louis, MO 63103. [www.sigmaaldrich.com](http://www.sigmaaldrich.com)  
Perkin Elmer, 940 Winter Street, Waltham, MA 02451. [www.perkinelmer.com](http://www.perkinelmer.com)

#### **Biochemistry Analyzer – Lactic acid**

Aliquot of extract analyzed for L-Lactate using YSI 2700 SELECT Biochemistry Analyzer equipped with an L-Lactate membrane. YSI User's Manual, page 4-7.

Samples injected into sample chamber of YSI Analyzer where L-Lactate diffuses into a membrane containing L-Lactate oxidase. The L-Lactate is immediately oxidized to hydrogen peroxide and pyruvate. The hydrogen peroxide is detected amperometrically at the platinum electrode surface. The current flow at the electrode is directly proportional to the hydrogen peroxide concentration, and hence to the L-Lactate concentration. Total lactic acid is determined by multiplying L-Lactate by 2.0.

YSI Incorporated Life Sciences, 1725 Brannum Lane, Yellow Springs, Ohio 45387. [www.ysilifescience.com](http://www.ysilifescience.com)